



# Anticancer activity and chemoprevention of xenobiotic organosulfurs in preclinical model systems

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## Abstract

There seems to be little doubt that xenobiotic and plant derived organosulfur compounds have enormous benefits for *in vitro* cellular functions and for a multitude of diseases, including cancer. Since there are numerous reviews on anticancer activities of plant organosulfurs, the focus herein will be on alterations associated with xenobiotic organosulfurs. Benefits of 2-mercaptoethanol (2-Me), N-Acetyl-cysteine, cysteamine, thioproline, piroxicam, disulfiram, amifostine, sulindac, celecoxib, oltipraz and their derivatives on transplanted homologous tumors and on autochthonous cancers with a viral-, radiation-, chemical carcinogen-, and undefined-etiology are assessed. Because all organosulfurs were not tested for activity in each of the etiology categories, comparative evaluations are restricted. In general, all 'appeared' to lower the incidence of cancer irrespective of etiology; however, since most of these values were determined at ages much younger than at a natural-end-of-life-age, differences most likely, instead, reflect a delayed initiation and/or a slowed progression of tumorigenesis. The poorest, long-term benefits of early intervention protocols occurred for viral- and chemical carcinogen-induced cancers. In addition, once tumorigenesis was beyond the initiation stage, outcomes of organosulfur therapies were extremely poor, indicating that they will not be of significant value as stand alone treatments. More importantly, except for the lifetime prevention of spontaneous and radiation-induced mammary tumors by daily dietary 2-Me, similar life long prevention of tumorigenesis was not achieved with other xenobiotics or any of nature's plant organosulfurs. These results raise an interesting question: Is the variability in incidence found for different organosulfurs associated with (a) their structure, (b) the length of the untreated latency period, (c) treatment duration/dose, and/or (d) the etiology-inducing agent?

**Keywords:** Xenobiotic organosulfurs, 2-mercaptoethanol, cancer, radiation, viral, chemical carcinogen, spontaneous, garlic, dietary

## Introduction

There seems to be little doubt that xenobiotic and plant-derived organosulfur compounds alter many biological processes that translate into enormous benefits for a multitude of diseases. Whereas the majority of investigations on food derived organosulfurs focused on anticarcinogenic bioactivity, research with xenobiotic organosulfurs focused on enhancement of *in vitro* immune functions. Surprisingly, even with the knowledge that immune functions play a major role in controlling cancer, there seems to be little cross collaboration or acknowledgment by the groups studying these different sources. This is especially disconcerting since in many respects the structural requirements for biological benefits of many of the food sulfur compounds/derivatives appear to be similar to, if not the same as, those postulated for bioactivity of xenobiotics [1-5].

Although there were a few early sporadic reports on organosulfur antioxidant benefits for virally induced and transplanted cancers [6-9], extensive investigations on the alteration of cellular events by organosulfurs began some 40 years ago when *in vitro*, cell mediated and humoral murine immune responses were shown to be dramatically enhanced by any of a multitude of structurally unrelated xenobiotic sulfhydryl compounds--2-mercaptoethanol (2-Me), dithiothreitol, reduced glutathione, and L-cysteine. Of these, the most effective was

2-Me, irrespective of whether it was added to protein-free or to autologous- or heterologous-sera supplemented culture media [10-14]. These findings led to an onslaught of reports defining benefits on immunological processes, and not surprisingly, on many other cell-types and processes (>1000 in PubMed). Regrettably, in many of these publications, the literature cited [15,16] as the origin of 2-Me's dramatic enhancement is totally incorrect. The cited reports replicated and confirmed our original research presented in 1971 at the First Congress of Immunology (workshop #71) in Washington, DC [17], findings that were at the time in press, and were published in early 1972 [10,11].

As might be expected from the extensive literature generated from the *in vitro* findings, investigations on 2-Me administered directly to animals soon followed (the 1980s). Makinodan and colleagues, the first to report on such benefits, demonstrated that the aged-associated decline in immune responsiveness both *in vitro* and *in situ* were corrected by either culturing with 2-Me or by a few (single or <5) injections of 2-Me [18-20]. A similar reversal of the age-decline of immune function in rats was subsequently reported [21,22]. Later, daily dietary exposure to 2-Me initiated at 16 weeks of age was shown to extend longevity [23], prevent both the decline of age-dependent, humoral and cell-mediated immune activity, curtail other aging processes associated with free-radical damage, and delay appearance

of spontaneous liver tumors [23,24]; the cancer findings supported earlier results obtained with a different xenobiotic organosulfur, cysteamine [25-27]. Soon thereafter, a colleague, Lee Wattenberg and his collaborators reported that the potent dithiolthione, antihelminthic sulfur drug, oltipraz, inhibited chemical carcinogen-induced neoplasia in mice [28]. Within a year, multiple compounds present in cruciferous vegetables, which previously had been demonstrated to inhibit chemical carcinogenesis, were identified to be organosulfurs, albeit with many different structures [29-35]. This in turn resulted in very active investigations (the 1990s) on the potential for controlling cancer by dietary plant organosulfurs. Interestingly, upon reflection on the sequence of progression over the past 40 years (history), it might be concluded that the enhancement that 2-Me imparted on *in vitro* immune functions initiated, directly or indirectly, an evolution of a new subject-area of research, namely bioactivity of organosulfurs on cellular and disease processes. Indeed, over the past decade, description of multitudes of other processes altered by xenobiotic, food, and complex organosulfur compounds has occurred—it seems there is no end to the discovery of new benefits. However, the present review will be limited to tumorigenic processes, with the focus on xenobiotics and long-term outcomes; benefits for other processes will be the subject of a later undertaking.

## Results and discussion

### Plant organosulfurs

Investigations on food organosulfurs and their selenium analogs suggested that they possess therapeutic value for multiple diseases; initially the most extensively studied was cancer [reviewed in 36,37]. Specifically, epidemiological data indicated that the incidence of stomach [38] and prostate cancer [39] was lower in populations that consumed large amounts of garlic. However, an evidence-based review [40] of the literature concluded that “only a remarkably few studies with generally small numbers of subjects were scientifically sound” and that only a “modest reduction in the risk of cancer was documented”. More convincing anti-cancer benefits were described with *in vitro* and rodent models for some of the organosulfurs or derivatives present in Brassica and Allium foods. The focus of the majority of these studies was on alterations of a variety of chemical carcinogen-induced specific tumorigenesis processes; *i.e.*, induction of phase 2 carcinogen-detoxifying enzymes, inhibition of phase 1 carcinogen-activating enzymes, cell cycle arrest, and apoptosis; the latter process being preferentially enhanced in cancer cells relative to normal cells [41]. Even though the degree of alteration of these processes correlated with tumor progression, survival was at most modestly extended, and indeed, cures or preventions were rarely, if ever, achieved [29-36,42-45]. This raises the question: Why the disparity? An answer may need to consider that few animal investigations were directed at alteration of normal anticancer surveillance processes (immune-mediated?); instead almost all used xenograft

transplant models and chemical carcinogens. In many respects, it could be argued that neither are hardly representative of normal cancer events. These type investigations further assumed that: (a) cancer caused by exposure to large doses of chemical carcinogens is a relevant model for tumorigenesis in humans; and (b) plant organosulfurs (i) can be consumed in sufficient quantities to be effective, (ii) are converted to bioactive forms *in situ*, and (iii) are not influenced by one another. Based on these uncertainties, it seems reasonable to ask: What are the realistic expectations of long-term, nutritional organosulfurs as preventive interventions? Evidence in the follow sections supports the conclusion that xenobiotics will be a more potent alternative.

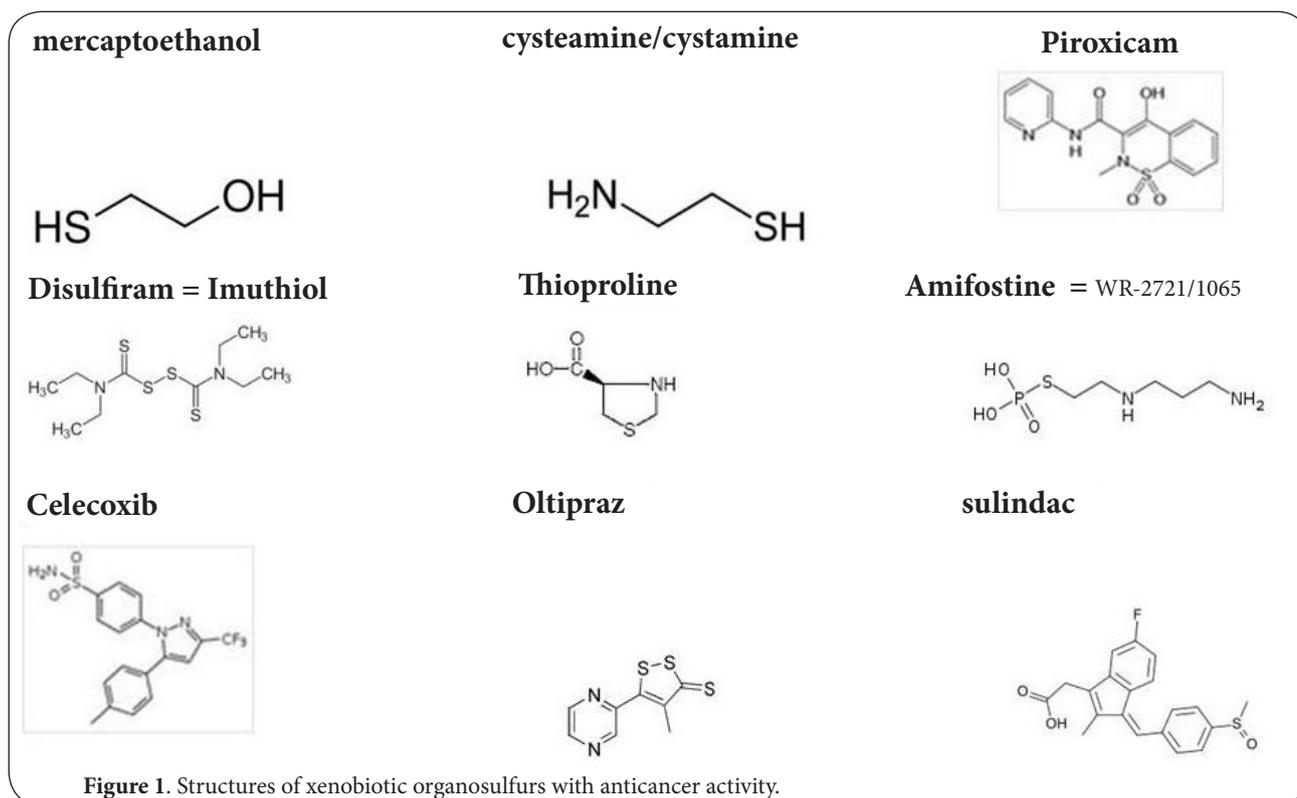
### Xenobiotic organosulfurs

The most extensive research with xenobiotic sulfur compounds originally focused on alterations of *in vitro* immunological processes; more recently benefits for a multitude of other cell-types, processes and diseases have been described [in preparation]. Since there is a lack of reviews on xenobiotic organosulfur bioactivities, the focus of this first report is limited to those that modify cancer processes (Figure 1). Each is assessed for alteration of transplanted autologous tumors and for autochthonous cancers with a viral-, radiation-, chemical carcinogen- or undefined-etiology (Table 1).

In many respects, bioactive organosulfurs from both sources share a number of similarities. Both are or can form ‘self’ or mixed disulfides, and those disulfides that are bioactive are structurally susceptible to enzymatic conversion to hypothesized H<sub>2</sub>S or sulfane sulfur [1-5,46-52]. And second, some of those not likely to form disulfide derivatives are still capable of generating H<sub>2</sub>S or sulfane sulfur. Arguments regarding these two potential end-products were recently reported [51,52]. However, other sulfur compounds (N-acetylcysteine, lipoic acid, glutathione, D-penicillamine, either as sulfhydryls or disulfides) have disease-altering capabilities, including cancer [53-57] and yet evidence that supports their generation of H<sub>2</sub>S or sulfane sulfur is lacking [1-5]. Irrespective of how xenobiotic or nature’s organosulfur compounds prevent development of, slow progression of, or are therapeutic post-occurrence of cancer (or any other disease), they have been categorized, directly or indirectly, as: (a) acting as free radical inhibitors/scavengers [58], (b) regulating gene expression [41,42,46,48-50], (c) maintaining critical allosteric disulfide configurations of cytoplasmic/membrane proteins [59,60], and/or (d) maintaining nature’s endogenous thiols—glutathione and thioredoxin—at an optimal redox balance for cellular functions [57,61,62]. Which, if any, of these categories best encompass clinical outcomes of structurally unique organosulfurs on different tumor-inducing insults remain to be defined; *in situ* mechanisms remain unresolved!

### Viral etiology

Murine strains infected with exogenous MMTV or endogenous



MuLV [66] develop mammary tumors and leukemia respectively. Early investigations by Harman described the effects of dietary supplementation of four sulfur antioxidants, 2-Me, cysteamine, cystamine, and L-cysteine [6,25] with C3H/J virgin females and AKR/J males. Since these experiments were done prior to Jackson Labs deriving MMTV(S)-free C3H by foster nursing, mammary tumors were presumably induced by milk-borne virus transferred during nursing. Each of the four test compounds were incorporated into feed. One was fed once a day as a powder, which it was noted was the time the animals ate the most, and a second was fed as a pellet, which was supplied ad libitum—both test-diets were started at weaning. Obviously, these two methods of delivery would result in different and unequal intakes over a 24 hour period as well as possible alterations during pellet-formulation. The incidence of mammary tumors was not altered and there was no significant increase in longevity of C3H fed the powder-diet containing 0.5% or 1.0% cysteamine, cystamine, or cysteine; the slightly extended longevity found with 0.5% 2-Me (estimated at 20,000 ug/day) was not statistically significant. In contrast, of the compounds tested in the pellet-diet, cysteamine at 1% (estimated intake of 40 mg/day) was the only one that increased median survival from 14.5 to 18.3 months (26%)—the importance of the diet formulation on survival by cysteamine was not addressed. In addition, this extension was accompanied by a delay in the onset and a lower incidence of tumors over a normal lifetime (how much

was not indicated). In a similar viral model [64], 2-Me (daily intake of 2500-3500 ug/day) added to water at weaning of C3H.OL and C3H.OH (H-2 congenic with C3H/HeDiSn) male and multiparous females did not change the 100% tumor incidence induced by exogenous MMTV(S), but did: (a) extend the age at which tumors became palpable by 31% (median); (b) increase median longevity 47%; and (c) increase median longevity, post-tumor detection, 75%.

With strains that develop leukemia, Harman found a 20 percent increase in the median survival for AKR/J fed (started at weaning) cysteine (0.5% and 1.0%), cysteamine (1.0%), or cystamine (0.5%) added to the powder-diet [6,25]. There was no alteration by the latter two at 0.5% and 1.0% or by 0.5% 2-Me. It was stated that prolongation was not due to prevention of leukemia, although a slowing would seem a probable explanation for the increased longevity. With the pellet-diet, median survival was increased 14.5% by cysteine only. Cystamine was tested at only 1.0% and based on the powder-diet results, would not have been expected to be effective at this dose. Interestingly, the non-sulfur, hydroxylamine in the pelleted-diet increased survival 8.3% at 1% and, 17.0% at 2%. With a different leukemic-prone strain, AKR/Cum, alteration of survival by 2-Me depended upon both dose and age at which treatment was initiated (Click, unpublished). At low doses (<400 ug/day), longevity was shortened 13.7% and 15.4% from medians of 306 and 273 days for untreated males and females, respectively. In contrast, increase in longevity

**Table 1. Benefits of individual organosulfurs--etiology agents, tumor types, and references.**

Etiologic agent	Sulfur compound	Tumor	Animal strain	Benefit <sup>1</sup>	Reference
<b>Viral</b>	2-Me	mammary	C3H/J, C3H.OL	N & S/DP	6, 64
	2-Me	leukemia	AKR/J	N	6
	2-Me	leukemia	AKR/Cum	S/DP	Click, unpublished
	cysteamine	mammary	C3H/J	LP	25
	cysteamine	leukemia	AKR/J	S/DP	6
	cystamine	mammary	C3H/J	N	6, 25
	cystamine	leukemia	AKR/J	S/DP	6,
	L-cysteine	mammary	C3H/J	N	6, 25
	L-cysteine	leukemia	AKR/J	S/DP	6, 25
<b>Radiation</b>	2-Me	mammary	B10.A(4R)	LP	77
	amifostine	sarcoma	C3Hf/Kam	S/DP	70
	amifostine	mammary	rat	S/DP	73
	amifostine	multiple	(C57XBALB/c)F1	S/DP	71, 72
	cysteamine	mammary	rat	S/DP	73
	cysteamine	intestinal	rat	S/DP	74
	piroxicam	colon	rat	S/DP	75
	Combination NAC + LA	lymphoma & rare	CBA/Ca	S/DP	76
	<b>Chemical carcinogen</b>	cysteamine	mammary	rat	S/DP
cysteamine		gastric	rat	S/DP	98
cysteamine		colon	rat	S/DP	99
cysteamine		liver	rat	S/DP	100
disulfiram		mammary	rat	N & S/DP	28, 83
disulfiram		forestomach	Ha/ICR	NLP	28
disulfiram		lung	A/HeJ	N	28
disulfiram		intestinal	CF1	NLP	84
disulfiram		liver	rat	S/DP	85, 86
disulfiram		bladder	rat	N & S/DP	86, 87
disulfiram		esophageal	rat	N	86
oltipraz		pulmonary & forestomach	ICR/Ha	S/DP	103
oltipraz		bladder	C57BL	S/DP	105
oltipraz		liver	rat	S/DP	106, 107
oltipraz		colon, breast, stomach, skin	rat	S/DP	104
piroxicam		intestinal	rat	N & S/DP	113-116
piroxicam		many	canine	S/DP	127
celecoxib		intestinal, mammary	rat	N & S/DP	117-120
sulindac		colon	human	S/DP	129, 130
sulindac		intestinal	C57min	PP	131, 132
sulindac	colon	Swiss	S/DP	134	
sulindac	colon	rat	S/DP	135, 136, 138-140	
sulindac	mammary	rat	S/DP	137	
<b>Transplant</b>	cysteamine	MC sarcoma	C57BL/P	S/DP & LP	27
	SH-blocked cysteamine	MC sarcoma	C57BL/P	N	27
	NH blocked cysteamine	MC sarcoma	C57BL/P	N	27
	cystamine	MC sarcoma	C57BL/P	N	27
	amifostine	MC sarcoma	C57BL/P	S/DP	27
	cysteamine	BAC/P	C3H/HeJ	S/DP	27
	cysteamine	sarcoma 1	A/J	S/DP	27
	cysteamine	Krebs-2	Swiss	S/DP & LP	27
	cysteamine	Ehrlich	Any strain	N	141
	2-Me disulfide	MC sarcoma	C57BL/P	enhanced	27
	thioglycerol	BAC/P	C3H/HeJ	N	27
	amifostine	BAC/P	C3H/HeJ	S/DP	27
	amifostine	sarcoma 1	A/J	S/DP	27
	amifostine	Krebs-2	Swiss	S/DP	27
	NAC	fibrosarcoma	C3H/He	N	55, 144

**Table 1. Continuation.**

Etiologic agent	Sulfur compound	Tumor	Animal strain	Benefit <sup>1</sup>	Reference
Spontaneous	2-Me	mammary	BXSB-Yaa+	LP	64
	2-Me	liver	(C57xC3H)F1	S/DP	23
	2-Me	liver/Dunn	CBA/Ca	S/DP	24, 151
	amifostine	multiple	(C57xBALB/c)F1	N	71, 72
	NAC	T-cell lymphoma	129Sv/C57B6Atm-	S/DP	146
	thioprolone	esophageal	rat	S/DP	148
	sulindac	intestinal	C57min	NLP	149, 150

<sup>1</sup>N = none, S/DP = slowed/delayed progression, PP = <100% incidence (partial prevention), LP = lifetime, 100% prevention, NLP = non-lifetime, 100% prevention.

was directly associated with the age at which a high dose ( $\geq 3500$   $\mu\text{g}/\text{day}$ ) was initiated---the later the start-age (in utero, 35 days, 150 days), the more longevity was increased (8.6%, 22.8%, 36.3%). Leukemia status was not determined, but was likely curtailed by the most favorable 150 day start age; # surviving >365 days increased from 28.6% to 75% for males and from 12.5% to 50% for females. In summary, the impact of initiating organosulfur treatment at weaning on two naturally-occurring, viral-induced cancers was quite poor. In addition, effectiveness depended upon which drug was used, the tumor model, and the dose. The largest increase in longevity of the leukemia-prone strain by 2-Me was when treatment was initiated late in life, which will become apparent, is opposite that for tumors caused by other etiology agents.

### Radiation etiology

It has been known for many years that numerous factors influence radiation carcinogenesis in animals. Agents that enhance or suppress these processes were recently reviewed [65]. It is also known that many types of damage caused by radiation can be ameliorated by antioxidants [66], including some exotic botanicals [67]. Those containing sulfur are especially effective and will be the only ones considered herein.

Among radio-protective agents for humans, WR-2721 (amifostine) is one of the most commonly used [68,69]. This prodrug, after *in situ* dephosphorylation, is a potent sulfhydryl-containing protector against early and late radiation damage of most normal tissues. One of the earliest antitumor studies was undertaken with C3Hf/Kam mice that received a single localized gamma-ray dose of 34 to 57 Gy directed to a syngeneic methylcholanthrene-induced fibrosarcoma previously transplanted into one of the hind limbs [70]. Thirty minutes prior to radiation, ca. 12 mg WR-2721 was or was not injected intraperitoneally. Those cured of the transplanted tumor were then observed for up to 786 days post-irradiation for histologically-different tumors. Recurrence in both untreated and drug treated animals occurred around 300 days. Thereafter, the rate of development was slower in the treated group. At termination of the experiment, the incidence in treated mice was 26% and that in controls was 87%. Using an identical scheme of drug treatment, (C57BL/6 x BALB/c) F<sub>1</sub>

male and female mice were radiated (total body) with 10 cGy of neutrons [71]. Cancers of connective and epithelial tissue origins were identified at necropsy as animals succumbed from natural causes (mostly cancer). Surprisingly, the incidence and mortality due to spontaneous tumors in non-irradiated, WR-2721 treated and not treated, males or females was not significantly different. Secondly, radiation shortened the lifespan due to neoplasia-related deaths. Thirdly, the age of tumor-linked death was sex associated---for females, radiation-shortened longevity was altered only in the first half of their lives, whereas for males, alteration occurred only in the second half. In addition, there was no difference in the incidence of tumors in radiated vs. nonirradiated animals for either sex indicating that radiation simply shortened the latency. Because the shift was so minimal, the unanswered question is: was the latency of normal, spontaneously occurring cancer simply shortened or was a new cancer induced by radiation? Furthermore, the unaltered incidence of spontaneous arising tumors by a single WR-2721 injection raises the question as to its value; a question reinforced by the mere 65 day extended survival of WR-2721-injected animals that were radiated with a higher dose (206 cGy) compared to similarly irradiated, non-WR-2721 treated controls [72].

Anticancer activities of WR-2721 and cysteamine were compared in pregnant rats in which palpable mammary tumors were induced by sublethal irradiation [73]. All rats were implanted with the tumor promoter diethylstilbestrol a month after termination of nursing. No spontaneous tumors developed in those not irradiated, an obvious advantage for interpretations. Nontreated controls radiated with 1.5 or 2.6 Gy had tumor incidences at termination (one year of age) of 71.4% and 92.3%, respectively. A single injection of WR-2721 or cysteamine 30 minutes prior to the 1.5 Gy dose significantly lowered the incidence to 23.8% and 20.8%, respectively; a reduction that, in part, may have been a consequence of an extended latency. Tumor prevention by either drug was less effective at the higher radiation dose.

In a different series of investigations, the influence of cysteamine was compared to ranitidine on intestinal metaplasia induced by irradiation of male rats. At the age of 5 weeks, the animals were locally radiated in the gastric region

with 10 Gy of X-rays at 3-day intervals for a total dose of 20 Gy [74]. After irradiation, the rats received either ranitidine (0.02% in their diet) or cysteamine (0.1% in drinking water). Unfortunately, the start date and length of treatment given were not compatible with the age at which tumors were assessed—treatment was continued for 2 months after the animals were sacrificed! At 7 months, the incidence and number of metaplasia foci in those that were treated with cysteamine were significantly lower than that in nontreated controls, whereas in those given ranitidine, the incidence was higher than controls. Thus, two organosulfurs sharing the cysteamine backbone (HS-C-C-NH<sub>2</sub> and R-C-S-C-C-NH-R') resulted in opposite outcomes for radiation-induced tumor formation.

Nonsteroidal antiinflammatory agents also altered neoplasia induced by radiation. Wistar female rats treated orally with 8.0 mg/kg piroxicam 30 minutes prior and 24 hours after localized pelvic 2250 cGy were found to have a significantly decreased incidence of, as well as a delay in, endoscopically detectable colonic cancer (primarily adenocarcinomas). When the experiment was terminated, 15 of 19 (79%) animals not treated had cancer compared to only 8 of 20 (40%) treated. The first cancer detected in a control animal was at 15 weeks post-irradiation compared to 36 weeks for a treated animal [75], suggesting that the lower incidence at one year post-irradiation was most likely a consequence of a delayed/slowed progression. In a more complex model [76], male CBA mice were fed a diet that, in part, included two organosulfurs, lipoic acid and N-acetyl-cysteine as part of a multi-antioxidant formulation. This diet reduced the risk of developing 0.5 Gy iron ion or 3 Gy proton-induced malignant lymphoma (>20%) and rare tumors (>10%) to almost that which occurred spontaneously (7% and 2.5%) over two years. Although it is not possible to access the importance of individual components, daily treatment with lipoic acid alone retarded progression of a xenograft-implanted, SkBr3 breast cancer cell line, strongly implicating the importance of a compound that occurs naturally as a sulfhydryl or disulfide [56].

In a more recent study [77], exposure of long-lived, B10.A(4R) mice to sublethal, 5.5 Gy ionizing gamma-rays at 288 days of age resulted in a 43% incidence of palpable mammary tumors over a normal lifetime. This incidence differed significantly from the 0% in (a) 44 non-irradiated controls that were or were not exposed to 2-Me (10<sup>-2</sup> M) drinking water (daily intake of 2500-3500 ug/m) starting at 90 days of age—2-Me treatment was terminated in half of these 24 hours after being radiated whereas the others were continued on treatment for their remaining lifetime, and (b) 50 irradiated that were treated continuously with 2-Me irrespective of whether the start days were at 90 day of age or 24 hours post radiation. An unexpected result of these studies was that irradiation significantly (P = 0.0002) shortened longevity 29% of animals pretreated with 2-Me from undefined causes (there was no obvious

signs indicative of cancer). This finding has relevance for the controversy on 'long term survival/safety' of currently used antioxidants as free radical scavengers in humans receiving radiotherapy [78,79] and the very recent concerns regarding the increased cancer incidence in pediatric patients due to the increased use of computed tomography [80].

In summary, a single or <5 injections of WR-2721, cysteamine, or piroxicam protected against radiation damage, but did not prevent development or progression of cancer. Delayed/slowed development is the most likely explanation for the lower incidences found, since in most studies, cancer was not determined over an entire normal lifespan. There may be a difference in outcomes after total body versus localized radiation—cancers induced by the latter appear to be more amenable to prevention by a single injection. A reasonable conclusion is that limited injections are not a practice that will successfully prevent radiation-induced cancer. Lastly, continuous exposure of mice to 2-Me, either only prior to, only post, or both prior to and post, sublethal total body irradiation prevented development of mammary tumors for an entire lifetime; no effort was made to examine for other types of tumors, although there were no obvious indications of any. An unexpected finding however indicated that there should be some caution in designing radiation protective protocols, since pretreatment with 2-Me for many weeks prior to total body radiation created a radiation-sensitive process that was manifested later in life by a shortened longevity. This has special relevance regarding 'long-term survival' of radiated patients that are 'protected' with various antioxidants [78,79], many of which possess potential active/activatable sulfur. Safe use of antioxidants for radiation protection needs to consider the: timing of antioxidant exposure relative to irradiation; structure of the antioxidant (sulfur vs. nonsulfur); duration of treatment; and the fact that sulfur containing antioxidants impact biological processes by means other than as free-radical scavengers [1-5,46-52,81,82].

### **Chemical carcinogen etiology Disulfiram and other dithiocarbamates**

Some of the earliest reports on organosulfur alterations of cancer were with disulfiram (200 mg/day), dimethyldithiocarbamate, and benzyl thiocyanate (cystine was ineffective). When added to the diet of 6 week old female Sprague-Dawley rats one week prior to oral intubation of 12 mg 7,12-dimethylbenz[a]-anthracene (DMBA), the 59-79% (three separate experiments) incidence of mammary tumors in untreated animals was reduced to 8-22%, 33%, and 8% by the three antioxidants at 23 weeks of age respectively; the number of tumors was also reduced. Further, a single, oral delivery of disulfiram at 24 hours before DMBA reduced the incidence from 59% to 11% [28]. In separate studies, disulfiram added to the diet (intake of 180 mg/day) for one week prior to and during carcinogen exposure, reduced the incidence of N-2-fluorenylacetylamide-induced (2-FAA) mammary tumors by 50% and extended the

mean latency period from 5 to 10 months; there was no effect on mammary tumors induced by the derivative, N-hydroxy-N-2-fluorenylacetylamine (N-OH-2-FAA) [83].

Even though disulfiram was approved and used as a common treatment of alcoholism, after the late 70's-early 80's, interest as an anticancer drug waned. It resurfaced within 10-15 years as a means to control chemical carcinogenic-induced tumors in rodents. It: (a) completely prevented the occurrence of benzo[a]pyrene (BP) induced tumors in the fore-stomach of Ha/ICR female mice terminated at 29 weeks but did not alter induction of pulmonary adenoma formation in A/HeJ female mice at 31 weeks [28]; (b) completely prevented 1,2-dimethylhydrazine- (DMH-) induced tumors of the large intestine of CF1 mice at 36 weeks [84]; (c) reduced liver tumors (66% and 95%) induced in rats by either diethylnitrosamine (DENA) or dimethylnitrosamine (DMNA) [85]; (d) prevented liver tumors, had no effect on esophageal and urinary bladder tumors, and actually increased lung tumors from 0% to 30% in rats, all induced by N-nitrosodibutylamine (NDBA) [86], and (e) reduced urinary bladder cancer from 100% to 13% in rats given N-n-butyl-N(4-hydroxybutyl)nitrosamine (BHBN) [87]. Surprisingly, when co-administrated with (a) DMNA, cell carcinomas of the paranasal sinus that were not found when either was given alone increased [88], and (b) NDBA, formation of lung tumors not normally induced by either individually were found [86]. It should be stressed that none of these treatments were followed for an entire lifetime. In summary, disulfiram effectively delayed and slowed progression of tumorigenesis in different organs induced by any of a number of different carcinogens. For other organs it was ineffective and when given in combination with certain carcinogens, actually enhanced tumorigenesis. It should not be unexpected that in several clinical trials, the results have been disappointing (www.clinicaltrials.gov).

Current postulated mechanisms of disulfiram activity appear to be multi-faceted with a complete understanding yet to be forthcoming. Based on investigations with cells in culture, various findings include: (a) cellular proteasome functions are inhibited [88,89], (b) inhibition of DNA methyltransferase (DNMT-1)--an enzyme that via a thiol group enhances global 5 methyl-cytosine content--resulted in a reduced level of methylated cytosine and a re-expression of epigenetically silenced genes that led to a reduced growth of prostate cancer cells *in vitro* and to a 40% reduction of growth as a xenograft [90]; (c) diethyldithiocarbamate (the active moiety) increased oxidative stress via lowering the level of reduced glutathione. This lower level was associated with DNA fragmentation and cell death [91], results that were reversed by pre-treatment with N-acetyl-cysteine [92-94]; and (d) increased mitochondrial antioxidant enzymes that were found to decline as tumor specific miRNAs declined [95,96]--miRNA levels that were returned to normal by disulfiram. Furthermore, these new levels were accompanied by lower levels of antioxidant proteins even though there was no decline in their messenger RNAs.

## Cysteamine

Using the rat-DMBA mammary tumor model (15 mg/kg DMBA IV), cysteamine was tested for anticancer activity by IP injection at a dose of 150 mg/kg 20 min prior to and 5 and 24 hr after DMBA [26]. Tumor incidence at 4 months was 67% (12 of 18) for those receiving only DMBA and 26% (4 of 17) for those treated with cysteamine. The incidence increased with time in both groups and by 11 months the incidence and total number of tumors in the cysteamine-treated animals was the same as that in the 4 month, DMBA non-treated group. Thus, cysteamine delayed initiation and/or slowed progression, but did not prevent induction. In contrast to extended mammary tumor latency, induction of adrenal necrosis and lesions of the small intestinal epithelium was not altered [26].

In a separate study, the 80% rat-DMBA mammary tumor incidence was reduced to 50% by cysteamine and to 44% by the selenium disulfide analogue, selenocystamine [99]. In animals with tumors, there was no statistical difference in the total tumor yield (2.82 vs. 2.92/animal), even though the animals were treated for a much longer period; *i.e.*, the drugs were added to the diet 2 weeks prior to oral administration of DMBA at 8 weeks of age and continued for the entire 31-33 week duration of the experiment. Interestingly, the level of cysteamine required to obtain an inhibition comparable to that of the selenium disulfide analog was 500-750-fold higher. A more appropriate comparison of the analog would have been to cystamine (the sulfur disulfide of cysteamine) based on *in vitro* requirements [2] and *in situ* effects of cysteamine and cystamine [27].

Three other rat cancers in which cysteamine significantly reduced the incidence and number of tumors were gastric adenocarcinomas induced by N-methyl-N'-nitro-N-nitrosoguanidine (MNNG) [98], colon tumors induced by azoxymethane (AOM) [99], and hepatocarcinomas induced by N-nitrosomorpholine (NNM) [100]. In each model, alterations were postulated to be mediated by catecholamines, specifically norepinephrine [101]. Again, as with most chemical carcinogenic agents, benefits for long-term survival were not determined--instead animals were sacrificed for biochemical and molecular analyses at pre-old-age stages.

## Oltipraz and dithiolethione derivatives

Oltipraz, an antihelmintic (schistosomicide), and related dithiolethiones were found to possess anticancer activity. The primary metabolites of oltipraz generated by two major pathways common to various mammalian species are: "oxidative desulfuration of the thione, which does not seem to be metabolized further"; and second, "desulfuration, methylation, and cleavage of the dithiolethione ring disulfide bond, followed by cyclization of the resulting unstable intermediate into pyrrolopyrazines" [102], which can be metabolized to other nonfunctional oxidized forms. Some the earliest benefits of oltipraz were reported by Wattenberg and his collaborators [103]. They demonstrated that a single

oral dose administered 24 or 48 hours prior to BP, also given orally, reduced the number of carcinogen-induced pulmonary adenomas and tumors of the forestomach in female ICR/Ha mice. Formation of pulmonary adenomas induced by oral administration of diethylnitrosamine or uracil mustard carcinogens were also reduced by oltipraz when given orally 48 hours earlier, but not as pronounced as that found for BP. Others found it inhibited chemically induced carcinogenesis of bladder, colon, breast, stomach, and skin cancer models [104 and ref therein]. In studies with the urinary bladder-specific carcinogen N-nitrosobutyl(4-hydroxybutyl)amine (BBN), it effectively reduced the incidence of tumors in conventional C57BL/6 mice, but was ineffective in C57BL-*Nrf2*<sup>-/-</sup> mice [105]. These results indicate that the nuclear factor, erythroid derived-E2-related factor 2 (*Nrf2*) pathway and its downstream target genes are responsible for BBN detoxification via phase 2 enzymes leading to diminished carcinogenesis.

Based upon induced hepatic phase II enzyme activities *in vitro* and a reduction in presumptive preneoplastic lesions in an aflatoxin B1-induced hepatic tumorigenesis rat model [106], various dithiolethione derivatives were compared for their ability to alter enzymes and disease. For these experiments treatment was started 3 weeks prior to a two week carcinogenic exposure and the experiment was terminated after another 5 weeks. Dietary concentrations of oltipraz and 17 derivatives induced hepatic phase II enzyme activities *in vivo* as well as produced marked inhibition of tumorigenesis [107]. Functional analysis indicated that gene alterations were associated with glutathione metabolism and with the *Nrf2* pathway. Of these compounds, 15 produced greater induction of NAD(P)H:quinone reductase and 11 yielded greater induction of glutathione S-transferase than oltipraz. Nine of these, spanning a range of gene alterations, were further tested for their ability to prevent AFB1-induced tumors. Six were found considerably more effective than oltipraz; interestingly, the most potent was the parent compound, 3H-1,2-dithiole-3-thione (D3T). An inverse correlation was found for *in vivo* phase II enzyme induction and tumor chemoprevention, indicating that enhancement of carcinogen detoxification pathways was a major contributor in preventing AFB1-induced cancer. Three of these compounds were further tested for their ability to induce specific gene functions. The parent compound, D3T, and oltipraz induced 226 common, differentially expressed liver genes 24 hours after Fischer F344 male rats were given three single doses on alternative days [108]. In contrast, forty genes had distinct responses; in addition, the response efficacy to oltipraz was weaker than that to D3T. Further studies demonstrated a comparable inactivation of protein tyrosine phosphatases by D3T and oltipraz via covalent modification of cysteine residues at the active site. This inactivation could contribute to the activation of *Nrf2* via the ERK1/2 signaling pathway [109]. In addition, oltipraz inhibited the enzyme, phosphatase SHP2, which may underlie its antiangiogenic properties. Disappointedly, similar differences in mean levels

of rectal tissue and lymphocyte GSH and GST were not found during a 6 month, Phase I study with 26 patients that previously had undergone resected colon polyps or were first-degree female relatives of breast cancer patients [110].

### Nonsteroidal anti-inflammatory Drugs (NSAIDs)

Another group of agents with anticancer properties, nonsteroidal anti-inflammatory drugs (NSAIDs), both non-sulfur and sulfur, are approved for treatment of diseases in humans, primarily for pain and inflammation. Oxicams (Piroxicam) and Sulindac are nonselective inhibitors (inhibit both COX 1 and 2), whereas Coxibs (Celecoxib) are sulfonamides with activity specific for COX-2. COX-2 specific inhibitors are preferred because of less serious side effects in the GI tract. Since benefits of various NSAIDs are reviewed elsewhere [111,112], only those in which their sulfur component impart benefits are considered herein.

### Oxicams and coxibs

Male Sprague-Dawley rats fed a high dietary dose (130 to 195 ppm) of piroxicam had fewer methylazoxyethanol acetate (MAM)-, N-methylnitrosourea (MNU)-, or AOM-induced intestinal tumors [113-116]. In these studies piroxicam was fed 1 wk after initiation of or 1 wk before, during, and after carcinogen treatment. A significant reduction in the incidence of intestinal tumors and number of tumors/animal at 5 months occurred relative to that of controls. Similar results were found for AOM-induced colon tumors in F344 rats when any of four doses of piroxicam were initiated at either one or 13 wks post exposure to carcinogen. However, if started at 23 wk after carcinogen exposure, only the two high doses resulted in slightly lower incidences and number of adenomas/adenocarcinomas per tumor-bearing animal. Furthermore, piroxicam had no effect on the incidence of AOM-induced tumors of the duodenum, and had no consistent alteration of the incidence of ear duct tumors, primarily squamous cell carcinomas [115]. It was concluded based on the latter results, plus those described for male Sprague-Dawley rats exposed to MAMA [116], that “the spectrum of tumor types that are susceptible to piroxicam is not universal and it does not prevent or cure neoplasia”—it like most other sulfur chemicals simply delays initiation or slows progression. Similar reductions in the incidence of oral cancer in male F344 rats exposed to water-laced 4-Nitroquinoline 1-oxide (NQO) and breast cancer in female Sprague-Dawley rats gavaged with DMBA were found after piroxicam [117] or celecoxib [117-120] treatments. The experiments were terminated at 26 and 17 weeks respectively and prevention was not obtained. Just as found for miRNA alterations by disulfiram [95,96], down-regulated miRNA (#29c), a tumor suppressor, was restored by celecoxib in human gastric cancer cells [121]. Disappointedly, analysis of two sets of data from the National Health Insurance Research Database lead to the conclusion that two selective COX-2 inhibitors, celecoxib and rofecoxib, may at most, benefit 10% of colorectal cancer patients [122].

Based on the inhibition of chemically induced colon tumors, plus the curtailment of Ornithine Decarboxylase (ODC), an enzyme involved in polyamine biosynthesis, in rodents by indomethacin suggested that lowering prostaglandin activity with specific inhibitors may further reduce tumorigenesis [123]. This was tested with D,L- $\alpha$ -difluoromethylornithine (DFMO), a specific irreversible inhibitor of ODC. Test diets were started 1 week prior to the first dose of AOM, and the rats were sacrificed 26 weeks later. Those that received either 0.05% or 0.1% DFMO in the drinking water or a high dose of piroxicam (2.6 mg) developed significantly fewer intestinal tumors than in controls. A low dose of piroxicam (1.3 mg) had no effect; however when combined with the low dose of DFMO, each acting through a different mechanism, reduced tumor formation more than DFMO alone [124]. In a separate report, certain dose combinations of these two drugs resulted in complete prevention of tumors induced by AOM for 56 weeks [125]. Similar benefits were obtained with a different COX-2 specific inhibitor, C-phycoyanin, in combination with piroxicam in DMH-induced precancerous colon polyps [126].

Based on the positive results obtained with rodents, piroxicam alone and in combination with either Deracoxib, cisplatin, Mitoxantrone, doxorubicin, or surgery was evaluated in dogs for anticancer benefits for urinary bladder, inflammatory mammary carcinoma, and oral squamous cell carcinoma [127 and ref therein]. Tumor-free survival, overall survival and biological response rates were at most modest. The combination of piroxicam and surgery appeared to result in the best survival advantage.

### Sulindac

As a prodrug, it undergoes reversible oxidation/reduction to the sulfide metabolite, a potent inhibitor of prostaglandin (PG) production, or is irreversibly converted to the sulfone metabolite, which was originally described to lack pharmacological benefits [128]. Sulindac was effective at preventing intestinal tumors in familial adenomatous polyposis patients that inherit a mutant allele of the *Apc3* gene [129,130], and in inhibiting tumor formation in a mouse model (*Apc/Min*) in which an allele of the homologous mouse *Apc* gene was inactivated by a mutation [131,132]. Similar inhibition was reported for (a) survival of malignant glioma cells *in vitro* due to a consequence of activation of an endoplasmic reticulum stress response (ERSR) [133]; (b) initiation of colon tumors induced in conventional mice by exposure to DMH (it did not cause regression of established tumors) [134]; and (c) DMH- and AOM-induced colonic tumors in rats, again more effectively at the initiation than the progression stage [135,136].

Although sulindac sulfone lacked prostaglandin synthetase inhibitory activity [128], it was found to have cancer chemopreventive activity for MNU-induced mammary [137] and for AOM-induced colon tumors [138,139], although effectiveness depended upon the time of administration (incidence was reduced most effectively when treatment was during the

initiation and early post-initiation periods—only minimal alteration occurred during the promotion/progression stage [140]. Thus, both the sulfide and sulfone metabolic products of sulindac were most effective when given early; in utero was slightly more effective than if started at weaning [132] and mechanism of chemoprevention was, in part, independent of the prostaglandin pathway.

### Transplanted tumors

One of the initial studies [141] in which organosulfurs were tested for effects on transplanted tumors, found that a number of antioxidants, including cysteine and cysteamine were not effective at preventing growth of Ehrlich ascites. In retrospect, such a finding is what might be predicted since this tumor lacks tumor-specific transplantation antigens [142]. These experiments were followed by tests with DL-2-mercapto-3-hydroxypropanal [9], and a series of reports on antioxidant's alterations of spontaneously arising tumors that have a viral etiology [6,25], (see preceding Virus section) and of transplanted solid tumors that were maintained by passage in the strain of origin (except Krebs-2 tumor) [27]. The four tumors in the study were: (a) Krebs-2 in Swiss mice, (b) methylcholanthrene-induced MC sarcoma in C57BL/P mice, (c) sarcoma 1 in A/J mice, and (d) BAC/P, a spontaneous breast adenocarcinoma derived from C3H/HeJ. Different antioxidants were injected IP daily, starting either on the day of tumor inoculation, at 24 hours or 7 days post tumor transplant. Treatment was terminated on day 12 for C57BL/P, A/J, C3H/He and day 18 for Swiss since Krebs-2 tumors grew at a slower pace. Cysteamine and WR-2721 antioxidants merely slowed growth without achieving any tumor free A/J and C3H survivors. The best benefits were obtained with C57BL/P males, in which there were 'numerous' (the incidence was not given) tumor-free survivors at 5 months for the two treatments started within 24 hours of tumor transplant. A lower incidence was found for Swiss mice transplanted with Krebs-2 tumors. In addition to differences between the various tumor/strain models, all antioxidants were not equally therapeutic. This was investigated more extensively with the C57BL/P MC-tumor model. Cysteamine was effective in its native form, but was ineffective when either the amino or thiol group, or both, were blocked. Mercaptopropylamine, the 3-carbon homolog of cysteamine, was not as effective as cysteamine. Thioglycerol, one of the most potent *in vitro* enhancers of immune functions [143], had only negligible activity. Cystamine, the disulfide dimer of cysteamine was completely ineffective—a result just opposite that found for enhancement of proliferation of sulfur-dependent L1210 lymphoma cells in the presence of a required, exogenous source of diamine oxidase [2]. The most surprising result was that during a 12 day treatment with the disulfides of 2-Me or cysteine, tumor growth was enhanced 214 or 187%!!!!!! In a different tumor transplant model, NAC was found to be ineffective, unless combined with an adoptive IL-2/LAK cell protocol against transplanted

UV radiation-induced fibrosarcoma in C3H/HeN mice [144].

In assessing potential benefits and mechanisms of cancer-modulating agents for treatment of human cancer, ideal model systems would utilize 'normal' immunocompetent animals because autochthonous organ-related tumors are: (a) located in 'normal' anatomical sites; (b) characterized by cellular heterogeneity; and (c) more representative of human disease counterparts. Thus, even though this section on autochthonous transplanted tumors is included, investigations relating to xenograft-transplanted tumors will not be considered. Results obtained with this model have been extensively reported by others and are almost exclusively related to chemical carcinogenic processes, plus being an unnatural model, have yet to contribute information, outside carcinogenic molecular processes, pertinent to cancer prevention/cures.

### Spontaneous (undefined etiology)

Sulfur antioxidants reported to have benefits for cancer of undefined etiology include (WR-2721), N-acetyl-cysteine (NAC), Thioproline, sulindac, and 2-Me. In early studies with WR-2721, a single injection of 400 ug/gm body wt did not alter spontaneous tumor development in (C57BL/6 x BALB/c) F<sub>1</sub> mice even though there was a slightly extended latency of tumor development post radiation—discussed above [71,72]. Investigations with NAC were mainly focused on C57BL/6-*Atm*<sup>-/-</sup> (AT-mutated deficient) mice, a model of the human disorder ataxia telangiectasia (AT), in which both have abnormal humoral and cellular immune functions [145]. Treatment with NAC increased median survival of this strain from 50 to 68 weeks and reduced the incidence of thymic lymphoma two-fold [146], presumably due to a lowering of the enhanced oxidative stress created by an increase in reactive oxygen species (ROS) or to an abnormal response to ROS. This conclusion is based on a similar ROS alteration by the unrelated antioxidant Tempol, which doubled the lifespan of this strain by delaying the onset of thymic lymphoma [147].

In humans, reflux disease is associated with Barrett's esophagus which may lead to an increased risk for esophageal adenocarcinoma (EAC), in part due to reactive nitrogen species. Thioproline (TOP) was tested in a Wistar rat model of human duodeno-gastroesophageal reflux disease that was created by anastomosing the jejunum side-by-side to the esophagogastric junction which allowed retention of normal stomach function [148]. Treatment with this nitrite-trapping scavenger was started after surgery and continued for the duration of the experiment (70 weeks post op) by supplementation in the feed at 0.5%. Interestingly, it did not suppress the overexpression of inducible nitric oxide synthase (iNOS) and did not significantly alter the rare occurrence of esophageal squamous cell carcinoma (5.6% in controls and 7.7% in TOP treated). The mechanism suggested to explain the absence of EACs in TOP treated animals versus the 38.9% incidence in the control group was: TOP "inhibits not only the production of nitroso compounds by nitrite-reducing

bacteria but also reactive nitrogen species (RNS) such as nitric oxide (NO), peroxyxynitrite (ONOO<sup>-</sup>) and N-nitroso compounds derived from reflux of duodenal contents".

Another organosulfur investigated was sulindac and its two bioactive metabolites in the C57BL/6J-*Min*/+ (Min-mice) model of familial adenomatous polyposis. In the first study, sulindac resulted in both a reduced incidence (100% to 10%) and number of tumors per mouse (11.9 vs. 0.1) when assayed 70 days post treatment at 110 days of age [149]. In an essentially identical protocol, the sulfide derivative also [150] resulted in a reduced incidence (100% to 30%) and number of tumors per mouse (33.2 vs. 0.6). These reductions were accompanied by decreased levels of PGE<sub>2</sub> and increased enterocyte apoptosis. These results contrast to the ineffectiveness found for the sulfone derivative (100% incidence and nonsignificant lowering of tumors/mouse to 21.9 [150]. This failure differed from its positive alteration of carcinogen-induced rat mammary and intestinal tumors [137-139]. However, interpretations need to be made with caution because ineffectiveness occurred with a dose 2–10x lower (sub-threshold?) than that used for the positive benefits. Irrespective, the anti-tumor effects in *Apc*-deficient animals imparted by low doses of sulindac are mediated by the sulfide metabolite and the benefits correlate with suppression of prostaglandin synthesis.

And lastly, 3 independent laboratories described alteration of spontaneous occurring cancers via undefined etiology agents by daily continuous exposure to 2-Me [23,24,64,151]. In the first report [23], 16 week old (C57BL x C3H/Anf) F<sub>1</sub> male mice were fed a diet either free of or containing 0.25% (w/w) 2-Me (this calculates to a daily consumption of ca. 7,500 ugm or ≤200 ugm/gm body wt) for the remainder of their lives. The median survival time was significantly increased from 840 to 938 days (an increase of 11.7%), in part due to a slower development of tumors (primarily hepatomas) The average incidence calculated from 4-5 different ages (all after the peak-incidence ages of 424 and 665 days for nontreated and treated respectively) was reduced from 84% to 60% by 2-Me. Unfortunately, examination for the presence of tumors was terminated at 134 weeks, namely at essentially the median survival age. Other changes that occurred that may have a bearing on tumor development was 2-Me delayed the decline of immune functions and slowed the appearance of free radical induced intracellular lipid peroxidation products. It is important to note that neither of these latter processes were prevented, they were simply delayed. The second study, with a different strain of mice, CBA/Ca also found that diseases associated with old-age were slowed [24,151]. The mice consumed considerably less (8 ugm daily) from their water starting at 5 months of age and continued until they were terminated at 20 months (610 days). The incidence of liver carcinomas was reduced from 45% to 6% and Dunn sarcomas from 8 to 3%. Other changes that occurred were an increase in humoral functions to heterologous sheep RBC antigens and a decrease in the autoreactive humoral response

to homologous murine RBCs. In the most recent report [64] 2-Me completely prevented the 100% incidence of mammary tumors in untreated BXS<sup>B</sup>-*Yaa*<sup>+</sup>/*J* mice, which increased the median longevity to 954 days from 650. Initiation of treatment was begun at weaning (40 days) and continued for the entire lifetime by adding 2-Me to the drinking water. The average daily consumption ranged from 2800 to 3500  $\mu\text{g}$  or 60-80  $\mu\text{g}/\text{gm}$  body wt [57].

Since the only complete life-long prevention of cancer was achieved with 2-Me, it is intriguing to speculate on mechanisms and future benefits it might impart, IF the 'poison' stigma can be overcome. Realization of a treatment that positively alters (derails) any aspect of cancer will depend on (a) if it alters tumorigenic processes, (b) how to integrate it with endogenous (preventive) mechanisms of amelioration, and/or (c) how to best incorporate it as an adjuvant with other interventions, especially *ex vivo* preparation of dendritic cell (DC), TSTA specific CTL, or anti-CD1 LAK vaccines. Such an effort seems warranted based on mechanism that can be envisaged from reports that demonstrated:

- a. 2-Me is the most potent enhancer of lymphoid functions of all species both *in situ* and in cultures supplemented with autologous sera,
- b. 2-Me inactivates functions of rodent and human Treg cells (cells that are a major impediment to *in situ* anticancer immune functions [152-155],
- c. thiols enhance CD4, CD8, and LAK proliferation and functions, all thought to play significant roles in cancer control [59,62,156-161],
- d. NAC, a relatively weak organosulfur, enhanced IL-2/LAK adoptive anticancer therapy of transplanted autologous cancer—some mice were cured [144], and
- e. age-associated depressed functions of DCs (and T cells) were partially restored by the sulfur drug, pyrrolidine dithiocarbamate [162].

## Conclusions

Whether any of the many structurally unique organosulfur drugs discussed herein as well as those present in plant-foods alter tumorigenesis by similar or distinct mechanisms is a difficult question that remains to be answered? Much of the uncertainty is because there is not a single model system in which each organosulfur was tested for anticancer activity. Thus, even though comparisons are severely restricted, a few generalized comments can be made. First, all organosulfurs 'appear' to lower the incidence of cancer; however, these values were invariably determined at specified ages much younger than a 'natural, end-of-life-age'. Thus, it is more accurate to view them as a consequence of tumor initiation being delayed and/or progression being slowed. Other generalizations are

- a. reduced and oxidized forms (cysteamine/cystamine) may result in different anticancer benefits—results that are not always consistent with *in vitro* activities [1,2],
- b. continuous vs. single exposure, as well as initiation of

- treatment prior to or soon after the inducing-event, resulted in the longest prolongation of latency,
- c. anticancer benefits were not the same for all etiology agents, and in some cases benefits were even organ specific,
- d. most drugs were effective against any of a number of different chemical carcinogens,
- e. combination with other drugs may have an enhanced or detrimental anticancer benefit,
- f. essentially all were not very effective against the two tumor inducing insults, chemical carcinogens and viruses, and
- g. slowing or preventing initiation was more easily achieved than altering established tumors,

Based on these generalizations it is almost certain that many of the anticancer benefits of xenobiotic organosulfurs are by multiple mechanisms, just as was postulated for the many uniquely structured food organosulfurs [163]. The most recent hypothesis involve gene control via alteration of specific miRNAs; supportive evidence is the differential gene activation by different structured organosulfurs [107-109,164].

Finally, perhaps the most important conclusion from all the models summarized herein is that organosulfurs will likely not be of value as stand alone cancer treatments, but may have value if combined with other therapies. The different degrees of delayed/slowed progression vs. complete prevention do raise two important points. First, it reinforces the concept that prevention is far more achievable than curative treatments. And second: Is the variability in incidence found for different organosulfurs associated with (a) their structure, (b) the length of the untreated latency period, (c) treatment duration/dose, and/or (d) the etiology-inducing agent? An answer should be extremely valuable in defining organosulfur cancer modalities that will yield better outcomes than those presently being found in clinical trials (www.clinicaltrials.gov).

## Competing interests

The author declares no competing interests.

## Author contributions

All aspects of the manuscript were contributed by the author.

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## References

1. Broome JD and Jeng MW. Promotion of replication in lymphoid cells

- by specific thiols and disulfides in vitro. Effects on mouse lymphoma cells in comparison with splenic lymphocytes. *J Exp Med.* 1973; **138**:574-92. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
2. Toohey JJ. Persulfide sulfur is a growth factor for cells defective in sulfur metabolism. *Biochem Cell Biol.* 1986; **64**:758-65. | [Article](#) | [PubMed](#)
  3. Cooper AJ and Pinto JT. Aminotransferase, L-amino acid oxidase and beta-lyase reactions involving L-cysteine S-conjugates found in allium extracts. Relevance to biological activity? *Biochem Pharmacol.* 2005; **69**:209-20. | [Article](#) | [PubMed](#)
  4. Cooper AJ and Pinto JT. Cysteine S-conjugate beta-lyases. *Amino Acids.* 2006; **30**:1-15. | [Article](#) | [PubMed](#)
  5. Cooper AJ, Krasnikov BF, Niatsetskaya ZV, Pinto JT, Callery PS, Villar MT, Artigues A and Bruschi SA. Cysteine S-conjugate beta-lyases: important roles in the metabolism of naturally occurring sulfur and selenium-containing compounds, xenobiotics and anticancer agents. *Amino Acids.* 2011; **41**:7-27. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
  6. Harman D. Prolongation of the normal life span by radiation protection chemicals. *J Gerontol.* 1957; **12**:257-63. | [Article](#) | [PubMed](#)
  7. Weisberger AS and Pinsky J. Tumor inhibition by a sulfhydryl-blocking agent related to an active principle of garlic (*Allium sativum*). *Cancer Res.* 1958; **18**:1301-8. | [Article](#) | [PubMed](#)
  8. Kroening F. Garlic as an Inhibitor for Spontaneous Tumors in Mice. *Acta Unio Int Contra Cancrum.* 1964; **20**:855-6. | [PubMed](#)
  9. Apple MA and Greenberg DM. Inhibitory effect of DL-2-mercapto-3-hydroxypropanol on growth of transplantable cancers in mice. *Cancer Chemother Rep.* 1969; **53**:195-8. | [PubMed](#)
  10. Click RE, Benck L and Alter BJ. Enhancement of antibody synthesis in vitro by mercaptoethanol. *Cell Immunol.* 1972; **3**:156-60. | [Article](#) | [PubMed](#)
  11. Click RE, Benck L and Alter BJ. Immune responses in vitro. I. Culture conditions for antibody synthesis. *Cell Immunol.* 1972; **3**:264-76. | [Article](#) | [PubMed](#)
  12. Heber-Katz E and Click RE. Immune responses in vitro. V. Role of mercaptoethanol in the mixed-leukocyte reaction. *Cell Immunol.* 1972; **5**:410-8. | [PubMed](#)
  13. Katz-Heber E, Peck AB and Click RE. Immune responses in vitro. II. Mixed leukocyte interaction in a protein-free medium. *Eur J Immunol.* 1973; **3**:379-85. | [Article](#) | [PubMed](#)
  14. Peck AB, Katz-Heber E and Click RE. Immune responses in vitro. IV. A comparison of the protein-free and mouse serum-supplemented mouse mixed lymphocyte interaction assays. *Eur J Immunol.* 1973; **3**:516-9. | [Article](#) | [PubMed](#)
  15. Chen C and Hirsch JG. Restoration of antibody-forming capacity in cultures of nonadherent spleen cells by mercaptoethanol. *Science.* 1972; **176**:60-1. | [Article](#) | [PubMed](#)
  16. Chen C and Hirsch JG. The effects of mercaptoethanol and of peritoneal macrophages on the antibody-forming capacity of nonadherent mouse spleen cells in vitro. *J Exp Med.* 1972; **136**:604-17. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
  17. First International Congress of Immunology—workshop #71—“Progress in Immunology”, ed., B Amos, Academic Press, New York and London, 1971.
  18. Makinodan T and Albright JW. Restoration of impaired immune functions in aging animals. II. Effect of mercaptoethanol in enhancing the reduced primary antibody responsiveness in vitro. *Mech Ageing Dev.* 1979; **10**:325-40. | [Article](#) | [PubMed](#)
  19. Makinodan T and Albright JW. Restoration of impaired immune functions in aging animals. III. Effect of mercaptoethanol in enhancing the reduced primary antibody responsiveness in vivo. *Mech Ageing Dev.* 1979; **11**:1-8. | [Article](#) | [PubMed](#)
  20. Chang MP, Tanaka JL, Stosic-Grujicic S, Yamamoto EK, Perkins EH, Strehler BL and Makinodan T. Restoration of impaired immune functions in aging animals. VI. Differential potentiating effect of 2-mercaptoethanol on young and old murine spleen cells. *Int J Immunopharmacol.* 1982; **4**:429-36. | [PubMed](#)
  21. Nauss KM, Connor AM and Newberne PM. Alterations in immune function in rats caused by dietary lipotrope deficiency: effect of culture medium, 2-mercaptoethanol and mitogen dose on the in vitro lymphocyte transformation response. *J Nutr.* 1982; **112**:2342-52. | [Article](#) | [PubMed](#)
  22. [Franklin RA, Li YM, Arkins S and Kelley KW. Glutathione augments in vitro proliferative responses of lymphocytes to concanavalin A to a greater degree in old than in young rats. *J Nutr.* 1990; **120**:1710-7. | [Article](#) | [PubMed](#)
  23. Heidrick ML, Hendricks LC and Cook DE. Effect of dietary 2-mercaptoethanol on the life span, immune system, tumor incidence and lipid peroxidation damage in spleen lymphocytes of aging BC3F1 mice. *Mech Ageing Dev.* 1984; **27**:341-58. | [Article](#) | [PubMed](#)
  24. Beregi E, Regius O, Rajczy K, Boross M and Penzes L. Effect of cigarette smoke and 2-mercaptoethanol administration on age-related alterations and immunological parameters. *Gerontology.* 1991; **37**:326-34. | [Article](#) | [PubMed](#)
  25. Harman D. Prolongation of the normal lifespan and inhibition of spontaneous cancer by antioxidants. *J Gerontol.* 1961; **16**:247-54. | [Article](#) | [PubMed](#)
  26. Marquardt H, Sapozink MD and Zedeck MS. Inhibition by cysteamine-HCl of oncogenesis induced by 7,12-dimethylbenz(alpha)anthracene without affecting toxicity. *Cancer Res.* 1974; **34**:3387-90. | [Article](#) | [PubMed](#)
  27. Apffel CA, Walker JE and Issarescu S. Tumor rejection in experimental animals treated with radioprotective thiols. *Cancer Res.* 1975; **35**:429-37. | [Article](#) | [PubMed](#)
  28. Wattenberg LW. Inhibition of carcinogenic and toxic effects of polycyclic hydrocarbons by several sulfur-containing compounds. *J Natl Cancer Inst.* 1974; **52**:1583-7. | [PubMed](#)
  29. Wargovich MJ. Diallyl sulfide, a flavonoid component of garlic (*Allium sativum*), inhibits dimethylhydrazine-induced colon cancer. *Carcinogenesis.* 1987; **8**:487-9. | [Article](#) | [PubMed](#)
  30. Hayes MA, Rushmore TH and Goldberg MT. Inhibition of hepatocarcinogenic responses to 1,2-dimethylhydrazine by diallyl sulfide, a component of garlic oil. *Carcinogenesis.* 1987; **8**:1155-7. | [Article](#) | [PubMed](#)
  31. Wattenberg LW. Inhibitory effects of benzyl isothiocyanate administered shortly before diethylnitrosamine or benzo[a]pyrene on pulmonary and forestomach neoplasia in A/J mice. *Carcinogenesis.* 1987; **8**:1971-3. | [Article](#) | [PubMed](#)
  32. Spornins VL, Barany G and Wattenberg LW. Effects of organosulfur compounds from garlic and onions on benzo[a]pyrene-induced neoplasia and glutathione S-transferase activity in the mouse. *Carcinogenesis.* 1988; **9**:131-4. | [Article](#) | [PubMed](#)
  33. Wargovich MJ, Woods C, Eng VW, Stephens LC and Gray K. Chemoprevention of N-nitrosomethylbenzylamine-induced esophageal cancer in rats by the naturally occurring thioether, diallyl sulfide. *Cancer Res.* 1988; **48**:6872-5. | [Article](#) | [PubMed](#)
  34. Wattenberg LW, Spornins VL and Barany G. Inhibition of N-nitrosodiethylamine carcinogenesis in mice by naturally occurring organosulfur compounds and monoterpenes. *Cancer Res.* 1989; **49**:2689-92. | [Article](#) | [PubMed](#)
  35. Liang D, Qin Y, Zhao W, Zhai X, Guo Z, Wang R, Tong L, Lin L, Chen H, Wong YC and Zhong Z. S-allylmercaptocysteine effectively inhibits the proliferation of colorectal cancer cells under in vitro and in vivo conditions. *Cancer Lett.* 2011; **310**:69-76. | [Article](#) | [PubMed](#)
  36. Wattenberg LW. Chemoprevention of cancer by naturally occurring and synthetic compounds. In: L. W. Wattenberg, M. Lipkin, C. W. Boone, and G. J. Kelloff (eds.), *Cancer Chemoprevent.* pp. 19-39. Boca Raton, FL: CRC Press, 1992.
  37. Milner JA. A historical perspective on garlic and cancer. *J Nutr.* 2001; **131**:1027S-31S. | [Article](#) | [PubMed](#)
  38. You WC, Blot WJ, Chang YS, Ershow A, Yang ZT, An Q, Henderson BE, Fraumeni JF, Jr. and Wang TG. Allium vegetables and reduced risk of

- stomach cancer. *J Natl Cancer Inst.* 1989; **81**:162-4. | [Article](#) | [PubMed](#)
39. Hsing AW, Chokkalingam AP, Gao YT, Madigan MP, Deng J, Gridley G and Fraumeni JF, Jr. **Allium vegetables and risk of prostate cancer: a population-based study.** *J Natl Cancer Inst.* 2002; **94**:1648-51. | [Article](#) | [PubMed](#)
40. Kim JY and Kwon O. **Garlic intake and cancer risk: an analysis using the Food and Drug Administration's evidence-based review system for the scientific evaluation of health claims.** *Am J Clin Nutr.* 2009; **89**:257-64. | [Article](#) | [PubMed](#)
41. Zeng H, Trujillo ON, Moyer MP and Botnen JH. **Prolonged sulforaphane treatment activates survival signaling in nontumorigenic NCM460 colon cells but apoptotic signaling in tumorigenic HCT116 colon cells.** *Nutr Cancer.* 2011; **63**:248-55. | [Article](#) | [PubMed](#)
42. Chandra-Kuntal K and Singh SV. **Diallyl trisulfide inhibits activation of signal transducer and activator of transcription 3 in prostate cancer cells in culture and in vivo.** *Cancer Prev Res (Phila).* 2010; **3**:1473-83. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
43. Ip C, Lisk DJ and Stoewsand GS. **Mammary cancer prevention by regular garlic and selenium-enriched garlic.** *Nutr Cancer.* 1992; **17**:279-86. | [Article](#) | [PubMed](#)
44. Conaway CC, Wang CX, Pittman B, Yang YM, Schwartz JE, Tian D, McIntee EJ, Hecht SS and Chung FL. **Phenethyl isothiocyanate and sulforaphane and their N-acetylcysteine conjugates inhibit malignant progression of lung adenomas induced by tobacco carcinogens in A/J mice.** *Cancer Res.* 2005; **65**:8548-57. | [Article](#) | [PubMed](#)
45. Wang X, Jiao F, Wang QW, Wang J, Yang K, Hu RR, Liu HC, Wang HY and Wang YS. **Aged black garlic extract induces inhibition of gastric cancer cell growth in vitro and in vivo.** *Mol Med Rep.* 2012; **5**:66-72. | [Article](#) | [PubMed](#)
46. Toohey JI. **Sulphane sulphur in biological systems: a possible regulatory role.** *Biochem J.* 1989; **264**:625-32. | [Pdf](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
47. Toohey JI. **Sulfur signaling: is the agent sulfide or sulfane?** *Anal Biochem.* 2011; **413**:1-7. | [Article](#) | [PubMed](#)
48. Wallace JL, Ferraz JG and Muscara MN. **Hydrogen sulfide: an endogenous mediator of resolution of inflammation and injury.** *Antioxid Redox Signal.* 2012; **17**:58-67. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
49. Miller TW, Wang EA, Gould S, Stein EV, Kaur S, Lim L, Amarnath S, Fowler DH and Roberts DD. **Hydrogen sulfide is an endogenous potentiator of T cell activation.** *J Biol Chem.* 2012; **287**:4211-21. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
50. Coletta C, Papapetropoulos A, Erdelyi K, Olah G, Modis K, Panopoulos P, Asimakopoulou A, Gero D, Sharina I, Martin E and Szabo C. **Hydrogen sulfide and nitric oxide are mutually dependent in the regulation of angiogenesis and endothelium-dependent vasorelaxation.** *Proc Natl Acad Sci U S A.* 2012; **109**:9161-6. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
51. Paul BD and Snyder SH. **H(2)S signalling through protein sulfhydration and beyond.** *Nat Rev Mol Cell Biol.* 2012; **13**:499-507. | [Article](#) | [PubMed](#)
52. Toohey JI. **The conversion of H(2)S to sulfane sulfur.** *Nat Rev Mol Cell Biol.* 2012; **13**:803. | [Article](#) | [PubMed](#)
53. Wu J, Levy EM and Black PH. **2-Mercaptoethanol and n-acetylcysteine enhance T cell colony formation in AIDS and ARC.** *Clin Exp Immunol.* 1989; **77**:7-10. | [PubMed Abstract](#) | [PubMed Full Text](#)
54. Herzenberg LA, De Rosa SC, Dubs JG, Roederer M, Anderson MT, Ela SW and Deresinski SC. **Glutathione deficiency is associated with impaired survival in HIV disease.** *Proc Natl Acad Sci U S A.* 1997; **94**:1967-72. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
55. Reliene R, Fleming SM, Chesselet MF and Schiestl RH. **Effects of antioxidants on cancer prevention and neuromotor performance in Atm deficient mice.** *Food Chem Toxicol.* 2008; **46**:1371-7. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
56. Feurecker B, Pirsig S, Seidl C, Aichler M, Feuchtinger A, Bruchelt G and Senekowitsch-Schmidtke R. **Lipoic acid inhibits cell proliferation of tumor cells in vitro and in vivo.** *Cancer Biol Ther.* 2012; **13**:1425-35. | [Article](#) | [PubMed](#)
57. Click RE. **Longevity of SLE-prone mice increased by dietary 2-mercaptoethanol via a mechanism imprinted within the first 28 days of life.** *Virulence.* 2010; **1**:516-22. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
58. Harman D. **Origin and evolution of the free radical theory of aging: a brief personal history, 1954-2009.** *Biogerontology.* 2009; **10**:773-81. | [Article](#) | [PubMed](#)
59. Edwards BS, Curry MS, Southon EA, Chong AS and Graf LH, Jr. **Evidence for a dithiol-activated signaling pathway in natural killer cell avidity regulation of leukocyte function antigen-1: structural requirements and relationship to phorbol ester- and CD16-triggered pathways.** *Blood.* 1995; **86**:2288-301. | [Article](#) | [PubMed](#)
60. Hogg PJ. **Targeting allosteric disulphide bonds in cancer.** *Nat Rev Cancer.* 2013; **13**:425-31. | [Article](#) | [PubMed](#)
61. Hiranruengchok R and Harris C. **Diamide-induced alterations of intracellular thiol status and the regulation of glucose metabolism in the developing rat conceptus in vitro.** *Teratology.* 1995; **52**:205-14. | [Article](#) | [PubMed](#)
62. Hadzic T, Li L, Cheng N, Walsh SA, Spitz DR and Knudson CM. **The role of low molecular weight thiols in T lymphocyte proliferation and IL-2 secretion.** *J Immunol.* 2005; **175**:7965-72. | [Article](#) | [PubMed](#)
63. Barnes RD and Brown K. **The lack of association of theta status and murine leukaemia virus content in the AKR.** *Br J Cancer.* 1975; **32**:678-9. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
64. Click RE. **Dietary supplemented 2-mercaptoethanol prevents spontaneous and delays virally-induced murine mammary tumorigenesis.** *Cancer Biol Ther.* 2013; **14**:521-6. | [Article](#) | [PubMed](#)
65. Kennedy AR. **Factors that modify radiation-induced carcinogenesis.** *Health Phys.* 2009; **97**:433-45. | [Article](#) | [PubMed](#)
66. Okunieff P, Swarts S, Keng P, Sun W, Wang W, Kim J, Yang S, Zhang H, Liu C, Williams JP, Huser AK and Zhang L. **Antioxidants reduce consequences of radiation exposure.** *Adv Exp Med Biol.* 2008; **614**:165-78. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
67. Sagar SM. **Can the therapeutic gain of radiotherapy be increased by concurrent administration of Asian botanicals?** *Integr Cancer Ther.* 2010; **9**:5-13. | [Article](#) | [PubMed](#)
68. Phillips TL. **Rationale for initial clinical trials and future development of radioprotectors.** *Cancer Clin Trials.* 1980; **3**:165-73. | [PubMed](#)
69. Yuhas JM, Spellman JM and Culo F. **The role of WR-2721 in radiotherapy and/or chemotherapy.** *Cancer Clin Trials.* 1980; **3**:211-6. | [PubMed](#)
70. Milas L, Hunter N, Stephens LC and Peters LJ. **Inhibition of radiation carcinogenesis in mice by S-2-(3-aminopropylamino)-ethylphosphorothioic acid.** *Cancer Res.* 1984; **44**:5567-9. | [Article](#) | [PubMed](#)
71. Carnes BA and Grdina DJ. **In vivo protection by the aminothiols WR-2721 against neutron-induced carcinogenesis.** *Int J Radiat Biol.* 1992; **61**:567-76. | [PubMed](#)
72. Grdina DJ, Carnes BA, Grahm D and Sigdestad CP. **Protection against late effects of radiation by S-2-(3-aminopropylamino)-ethylphosphorothioic acid.** *Cancer Res.* 1991; **51**:4125-30. | [Article](#) | [PubMed](#)
73. Inano H, Onoda M, Suzuki K, Kobayashi H and Wakabayashi K. **Inhibitory effects of WR-2721 and cysteamine on tumor initiation in mammary glands of pregnant rats by radiation.** *Radiat Res.* 2000; **153**:68-74. | [Article](#) | [PubMed](#)
74. Watanabe H, Kamikawa M, Nakagawa Y, Takahashi T and Ito A. **The effects of ranitidine and cysteamine on intestinal metaplasia induced by X-irradiation in rats.** *Acta Pathol Jpn.* 1988; **38**:1285-96. | [Article](#) | [PubMed](#)
75. Northway MG, Scobey MW, Cassidy KT and Geisinger KR. **Piroxicam decreases postirradiation colonic neoplasia in the rat.** *Cancer.* 1990; **66**:2300-5. | [Article](#) | [PubMed](#)
76. Kennedy AR, Davis JG, Carlton W and Ware JH. **Effects of dietary antioxidant supplementation on the development of malignant**

- lymphoma and other neoplastic lesions in mice exposed to proton or iron-ion radiation.** *Radiat Res.* 2008; **169**:615-25. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
77. Click RE: Dietary supplemented 2-mercaptoethanol alters in situ radiation-sensitive processes----potential cancer therapeutic or adjuvant and as an antioxidant protector. Submitted
78. Lawenda BD, Kelly KM, Ladas EJ, Sagar SM, Vickers A and Blumberg JB. **Should supplemental antioxidant administration be avoided during chemotherapy and radiation therapy?** *J Natl Cancer Inst.* 2008; **100**:773-83. | [Article](#) | [PubMed](#)
79. Bhutani M and Pathak AK. **Re: Should supplemental antioxidant administration be avoided during chemotherapy and radiation therapy?** *J Natl Cancer Inst.* 2008; **100**:1334. | [Article](#) | [PubMed](#)
80. Miglioretti DL, Johnson E, Williams A, Greenlee RT, Weinmann S, Solberg LI, Feigelson HS, Roblin D, Flynn MJ, Vanneman N and Smith-Bindman R. **The Use of Computed Tomography in Pediatrics and the Associated Radiation Exposure and Estimated Cancer Risk.** *JAMA Pediatr.* 2013; 1-8. | [Article](#) | [PubMed](#)
81. Biaglow JE, Varnes ME, Clark EP and Epp ER. **The role of thiols in cellular response to radiation and drugs.** *Radiat Res.* 1983; **95**:437-55 5. | [Article](#) | [PubMed](#)
82. Ratnam DV, Ankola DD, Bhardwaj V, Sahana DK and Kumar MN. **Role of antioxidants in prophylaxis and therapy: A pharmaceutical perspective.** *J Control Release.* 2006; **113**:189-207. | [Article](#) | [PubMed](#)
83. Malejka-Giganti D, McIver RC and Rydell RE. **Inhibitory effect of disulfiram on rat mammary tumor induction by N-2-fluorenylacetylamine and on its metabolic conversion to N-hydroxy-N-2-fluorenylacetylamine.** *J Natl Cancer Inst.* 1980; **64**:1471-7. | [PubMed](#)
84. Wattenberg LW. **Inhibition of dimethylhydrazine-induced neoplasia of the large intestine by disulfiram.** *J Natl Cancer Inst.* 1975; **54**:1005-6. | [PubMed](#)
85. Schmahl D, Kruger FW, Habs M and Diehl B. **Influence of disulfiram on the organotropy of the carcinogenic effect of dimethylnitrosamine and diethylnitrosamine in rats.** *Z Krebsforsch Klin Onkol Cancer Res Clin Oncol.* 1976; **85**:271-6. | [Article](#) | [PubMed](#)
86. Schweinsberg F, Weissenberger I and Burkle V. **The effect of disulfiram on the carcinogenicity of nitrosamines.** *IARC Sci Publ.* 1982; 649-57. | [PubMed](#)
87. Irving CC, Tice AJ and Murphy WM. **Inhibition of N-n-butyl-N-(4-hydroxybutyl)nitrosamine-induced urinary bladder cancer in rats by administration of disulfiram in the diet.** *Cancer Res.* 1979; **39**:3040-3. | [Article](#) | [PubMed](#)
88. Kona FR, Buac D and A MB. **Disulfiram, and disulfiram derivatives as novel potential anticancer drugs targeting the ubiquitin-proteasome system in both preclinical and clinical studies.** *Curr Cancer Drug Targets.* 2011; **11**:338-46. | [Article](#) | [PubMed](#)
89. Skrott Z and Cvek B. **Diethyldithiocarbamate complex with copper: the mechanism of action in cancer cells.** *Mini Rev Med Chem.* 2012; **12**:1184-92. | [Article](#) | [PubMed](#)
90. Lin J, Haffner MC, Zhang Y, Lee BH, Brennen WN, Britton J, Kachhap SK, Shim JS, Liu JO, Nelson WG, Yegnasubramanian S and Carducci MA. **Disulfiram is a DNA demethylating agent and inhibits prostate cancer cell growth.** *Prostate.* 2011; **71**:333-43. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
91. Burkitt MJ, Bishop HS, Milne L, Tsang SY, Provan GJ, Nobel CS, Orrenius S and Slater AF. **Dithiocarbamate toxicity toward thymocytes involves their copper-catalyzed conversion to thiuram disulfides, which oxidize glutathione in a redox cycle without the release of reactive oxygen species.** *Arch Biochem Biophys.* 1998; **353**:73-84. | [Article](#) | [PubMed](#)
92. Grosicka-Maciag E, Kurpios-Piec D, Grzela T, Czczot H, Skrzycki M, Szumilo M and Rahden-Staron I. **Protective effect of N-acetyl-L-cysteine against disulfiram-induced oxidative stress and apoptosis in V79 cells.** *Toxicol Appl Pharmacol.* 2010; **248**:210-6. | [Article](#) | [PubMed](#)
93. Rahden-Staron I, Grosicka-Maciag E, Kurpios-Piec D, Czczot H, Grzela T and Szumilo M. **The effects of sodium diethyldithiocarbamate in fibroblasts V79 cells in relation to cytotoxicity, antioxidative enzymes, glutathione, and apoptosis.** *Arch Toxicol.* 2012; **86**:1841-50. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
94. Conticello C, Martinetti D, Adamo L, Buccheri S, Giuffrida R, Parrinello N, Lombardo L, Anastasi G, Amato G, Cavalli M, Chiarenza A, De Maria R, Giustolisi R, Gulisano M and Di Raimondo F. **Disulfiram, an old drug with new potential therapeutic uses for human hematological malignancies.** *Int J Cancer.* 2012; **131**:2197-203. | [Article](#) | [PubMed](#)
95. Xu Y, Fang F, Zhang J, Josson S, St Clair WH and St Clair DK. **miR-17\* suppresses tumorigenicity of prostate cancer by inhibiting mitochondrial antioxidant enzymes.** *PLoS One.* 2010; **5**:e14356. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
96. Mutallip M, Nohata N, Hanazawa T, Kikkawa N, Horiguchi S, Fujimura L, Kawakami K, Chiyomaru T, Enokida H, Nakagawa M, Okamoto Y and Seki N. **Glutathione S-transferase P1 (GSTP1) suppresses cell apoptosis and its regulation by miR-133alpha in head and neck squamous cell carcinoma (HNSCC).** *Int J Mol Med.* 2011; **27**:345-52. | [Article](#) | [PubMed](#)
97. Ip C and Ganther HE. **Comparison of selenium and sulfur analogs in cancer prevention.** *Carcinogenesis.* 1992; **13**:1167-70. | [Article](#) | [PubMed](#)
98. Tatsuta M, Iishi H, Yamamura H, Baba M, Mikuni T and Taniguchi H. **Inhibitory effect of prolonged administration of cysteamine on experimental carcinogenesis in rat stomach induced by N-methyl-N'-nitro-N-nitrosoguanidine.** *Int J Cancer.* 1988; **41**:423-6. | [Article](#) | [PubMed](#)
99. Tatsuta M, Iishi H, Baba M and Taniguchi H. **Tissue norepinephrine depletion as a mechanism for cysteamine inhibition of colon carcinogenesis induced by azoxymethane in Wistar rats.** *Int J Cancer.* 1989; **44**:1008-11. | [Article](#) | [PubMed](#)
100. Tatsuta M, Iishi H and Baba M. **Inhibition by cysteamine of hepatocarcinogenesis induced by N-nitrosomorpholine in Sprague-Dawley rats.** *Int J Cancer.* 1989; **44**:529-33. | [Article](#) | [PubMed](#)
101. Tatsuta M, Iishi H, Baba M and Taniguchi H. **Attenuating effect of the monoamine oxidase inhibitor furazolidone on the anti-carcinogenic effect of cysteamine on gastric carcinogenesis induced by N-methyl-N'-nitro-N-nitrosoguanidine in Wistar rats.** *Int J Cancer.* 1991; **48**:605-8. | [Article](#) | [PubMed](#)
102. Bieder A, Decouvelaere B, Gaillard C, Depaire H, Heusse D, Ledoux C, Lemar M, Le Roy JP, Raynaud L, Snozzi C and et al. **Comparison of the metabolism of oltipraz in the mouse, rat and monkey and in man. Distribution of the metabolites in each species.** *Arzneimittelforschung.* 1983; **33**:1289-97. | [PubMed](#)
103. Wattenberg LW and Bueding E. **Inhibitory effects of 5-(2-pyrazinyl)-4-methyl-1,2-dithiol-3-thione (Oltipraz) on carcinogenesis induced by benzo[a]pyrene, diethylnitrosamine and uracil mustard.** *Carcinogenesis.* 1986; **7**:1379-81. | [Article](#) | [PubMed](#)
104. Kelloff GJ, Boone CW, Crowell JA, Steele VE, Lubet R and Sigman CC. **Chemopreventive drug development: perspectives and progress.** *Cancer Epidemiol Biomarkers Prev.* 1994; **3**:85-98. | [Article](#) | [PubMed](#)
105. Iida K, Itoh K, Kumagai Y, Oyasu R, Hattori K, Kawai K, Shimazui T, Akaza H and Yamamoto M. **Nrf2 is essential for the chemopreventive efficacy of oltipraz against urinary bladder carcinogenesis.** *Cancer Res.* 2004; **64**:6424-31. | [Article](#) | [PubMed](#)
106. Kensler TW, Groopman JD, Eaton DL, Curphey TJ and Roebuck BD. **Potent inhibition of aflatoxin-induced hepatic tumorigenesis by the monofunctional enzyme inducer 1,2-dithiole-3-thione.** *Carcinogenesis.* 1992; **13**:95-100. | [Article](#) | [PubMed](#)
107. Maxuitenko YY, Libby AH, Joyner HH, Curphey TJ, MacMillan DL, Kensler TW and Roebuck BD. **Identification of dithiolethiones with better chemopreventive properties than oltipraz.** *Carcinogenesis.* 1998; **19**:1609-15. | [Article](#) | [PubMed](#)
108. Tran QT, Xu L, Phan V, Goodwin SB, Rahman M, Jin VX, Sutter CH, Roebuck BD, Kensler TW, George EO and Sutter TR. **Chemical genomics of cancer chemopreventive dithiolethiones.** *Carcinogenesis.* 2009; **30**:480-6. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
109. Bhattacharyya S, Zhou H, Seiner DR and Gates KS. **Inactivation of protein tyrosine phosphatases by oltipraz and other cancer chemopreventive 1,2-dithiole-3-thiones.** *Bioorg Med Chem.* 2010;

- 18:5945-9. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
110. Benson AB, 3rd, Olopade OI, Ratain MJ, Rademaker A, Mobarhan S, Stucky-Marshall L, French S and Dolan ME. **Chronic daily low dose of 4-methyl-5-(2-pyrazinyl)-1,2-dithiole-3-thione (Oltipraz) in patients with previously resected colon polyps and first degree female relatives of breast cancer patients.** *Clin Cancer Res.* 2000; **6**:3870-7. | [Article](#) | [PubMed](#)
111. Thun MJ, Henley SJ and Patrono C. **Nonsteroidal anti-inflammatory drugs as anticancer agents: mechanistic, pharmacologic, and clinical issues.** *J Natl Cancer Inst.* 2002; **94**:252-66. | [Article](#) | [PubMed](#)
112. Rao CV and Reddy BS: **NSAIDs and chemoprevention.** *Current Cancer Drug Targets* 2004, **4**:29-42. | [Pdf](#)
113. Pollard M, Luckert PH and Schmidt MA. **The suppressive effect of piroxicam on autochthonous intestinal tumors in the rat.** *Cancer Lett.* 1983; **21**:57-61. | [Article](#) | [PubMed](#)
114. Pollard M and Luckert PH. **Effect of piroxicam on primary intestinal tumors induced in rats by N-methylnitrosourea.** *Cancer Lett.* 1984; **25**:117-21. | [Article](#) | [PubMed](#)
115. Reddy BS, Maruyama H and Kelloff G. **Dose-related inhibition of colon carcinogenesis by dietary piroxicam, a nonsteroidal antiinflammatory drug, during different stages of rat colon tumor development.** *Cancer Res.* 1987; **47**:5340-6. | [Article](#) | [PubMed](#)
116. Pollard M and Luckert PH. **Prevention and treatment of primary intestinal tumors in rats by piroxicam.** *Cancer Res.* 1989; **49**:6471-3. | [Article](#) | [PubMed](#)
117. McCormick DL, Phillips JM, Horn TL, Johnson WD, Steele VE and Lubet RA. **Overexpression of cyclooxygenase-2 in rat oral cancers and prevention of oral carcinogenesis in rats by selective and nonselective COX inhibitors.** *Cancer Prev Res (Phila).* 2010; **3**:73-81. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
118. Harris RE, Alshafie GA, Abou-Issa H and Seibert K. **Chemoprevention of breast cancer in rats by celecoxib, a cyclooxygenase 2 inhibitor.** *Cancer Res.* 2000; **60**:2101-3. | [Article](#) | [PubMed](#)
119. Abou-Issa HM, Alshafie GA, Seibert K, Koki AT, Masferrer JL and Harris RE. **Dose-response effects of the COX-2 inhibitor, celecoxib, on the chemoprevention of mammary carcinogenesis.** *Anticancer Res.* 2001; **21**:3425-32. | [PubMed](#)
120. Dai ZJ, Ma XB, Kang HF, Gao J, Min WL, Guan HT, Diao Y, Lu WF and Wang XJ. **Antitumor activity of the selective cyclooxygenase-2 inhibitor, celecoxib, on breast cancer in Vitro and in Vivo.** *Cancer Cell Int.* 2012; **12**:53. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
121. Saito Y, Suzuki H, Imaeda H, Matsuzaki J, Hirata K, Tsugawa H, Hibino S, Kanai Y, Saito H and Hibi T. **The tumor suppressor microRNA-29c is downregulated and restored by celecoxib in human gastric cancer cells.** *Int J Cancer.* 2013; **132**:1751-60. | [Article](#) | [PubMed](#)
122. Yang YH, Cheng CL, Ho PS and Ko YC. **The role of chemoprevention by selective cyclooxygenase-2 inhibitors in colorectal cancer patients - a population-based study.** *BMC Cancer.* 2012; **12**:582. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
123. Verma AK, Ashendel CL and Boutwell RK. **Inhibition by prostaglandin synthesis inhibitors of the induction of epidermal ornithine decarboxylase activity, the accumulation of prostaglandins, and tumor promotion caused by 12-O-tetradecanoylphorbol-13-acetate.** *Cancer Res.* 1980; **40**:308-15. | [Article](#) | [PubMed](#)
124. Nigro ND, Bull AW and Boyd ME. **Inhibition of intestinal carcinogenesis in rats: effect of difluoromethylornithine with piroxicam or fish oil.** *J Natl Cancer Inst.* 1986; **77**:1309-13. | [Article](#) | [PubMed](#)
125. Reddy BS, Nayini J, Tokumo K, Rigotty J, Zang E and Kelloff G. **Chemoprevention of colon carcinogenesis by concurrent administration of piroxicam, a nonsteroidal antiinflammatory drug with D,L-alpha-difluoromethylornithine, an ornithine decarboxylase inhibitor, in diet.** *Cancer Res.* 1990; **50**:2562-8. | [Article](#) | [PubMed](#) Abstract | [PubMed Full Text](#)
126. Saini MK, Vaiphei K and Sanyal SN. **Chemoprevention of DMH-induced rat colon carcinoma initiation by combination administration of piroxicam and C-phycocyanin.** *Mol Cell Biochem.* 2012; **361**:217-28. | [Article](#) | [PubMed](#)
127. Robat C, Burton J, Thamm D and Vail D. **Retrospective evaluation of doxorubicin-piroxicam combination for the treatment of transitional cell carcinoma in dogs.** *J Small Anim Pract.* 2013; **54**:67-74. | [Article](#) | [PubMed](#)
128. Duggan DE, Hooke KF, Risley EA, Shen TY and Arman CG. **Identification of the biologically active form of sulindac.** *J Pharmacol Exp Ther.* 1977; **201**:8-13. | [Article](#) | [PubMed](#)
129. Giardiello FM, Hamilton SR, Krush AJ, Piantadosi S, Hyland LM, Celano P, Booker SV, Robinson CR and Offerhaus GJ. **Treatment of colonic and rectal adenomas with sulindac in familial adenomatous polyposis.** *N Engl J Med.* 1993; **328**:1313-6. | [Article](#) | [PubMed](#)
130. Nugent KP, Farmer KC, Spigelman AD, Williams CB and Phillips RK. **Randomized controlled trial of the effect of sulindac on duodenal and rectal polyposis and cell proliferation in patients with familial adenomatous polyposis.** *Br J Surg.* 1993; **80**:1618-9. | [Article](#) | [PubMed](#)
131. Boolbol SK, Dannenberg AJ, Chadburn A, Martucci C, Guo XJ, Ramonetti JT, Abreu-Goris M, Newmark HL, Lipkin ML, DeCosse JJ and Bertagnolli MM. **Cyclooxygenase-2 overexpression and tumor formation are blocked by sulindac in a murine model of familial adenomatous polyposis.** *Cancer Res.* 1996; **56**:2556-60. | [Article](#) | [PubMed](#)
132. Beazer-Barclay Y, Levy DB, Moser AR, Dove WF, Hamilton SR, Vogelstein B and Kinzler KW. **Sulindac suppresses tumorigenesis in the Min mouse.** *Carcinogenesis.* 1996; **17**:1757-60. | [Article](#) | [PubMed](#)
133. White MC, Johnson GG, Zhang W, Hobrath JV, Piazza GA and Grimaldi M. **Sulindac sulfide inhibits sarcoendoplasmic reticulum Ca2+ ATPase, induces endoplasmic reticulum stress response, and exerts toxicity in glioma cells: relevant similarities to and important differences from celecoxib.** *J Neurosci Res.* 2013; **91**:393-406. | [Article](#) | [PubMed](#) Abstract | [PubMed Full Text](#)
134. Moorghen M, Ince P, Finney KJ, Sunter JP, Appleton DR and Watson AJ. **A protective effect of sulindac against chemically-induced primary colonic tumours in mice.** *J Pathol.* 1988; **156**:341-7. | [Article](#) | [PubMed](#)
135. Skinner SA, Penney AG and O'Brien PE. **Sulindac inhibits the rate of growth and appearance of colon tumors in the rat.** *Arch Surg.* 1991; **126**:1094-6. | [Article](#) | [PubMed](#)
136. Rao CV, Rivenson A, Simi B, Zang E, Kelloff G, Steele V and Reddy BS. **Chemoprevention of colon carcinogenesis by sulindac, a nonsteroidal anti-inflammatory agent.** *Cancer Res.* 1995; **55**:1464-72. | [Article](#) | [PubMed](#)
137. Thompson HJ, Jiang C, Lu J, Mehta RG, Piazza GA, Paranka NS, Pamukcu R and Ahnen DJ. **Sulfone metabolite of sulindac inhibits mammary carcinogenesis.** *Cancer Res.* 1997; **57**:267-71. | [Article](#) | [PubMed](#)
138. Alberts DS, Hixson L, Ahnen D, Bogert C, Einspahr J, Paranka N, Brendel K, Gross PH, Pamukcu R and Burt RW. **Do NSAIDs exert their colon cancer chemoprevention activities through the inhibition of mucosal prostaglandin synthetase?** *J Cell Biochem Suppl.* 1995; **22**:18-23. | [PubMed](#)
139. Piazza GA, Alberts DS, Hixson LJ, Paranka NS, Li H, Finn T, Bogert C, Guillen JM, Brendel K, Gross PH, Sperl G, Ritchie J, Burt RW, Ellsworth L, Ahnen DJ and Pamukcu R. **Sulindac sulfone inhibits azoxymethane-induced colon carcinogenesis in rats without reducing prostaglandin levels.** *Cancer Res.* 1997; **57**:2909-15. | [Article](#) | [PubMed](#)
140. Reddy BS, Kawamori T, Lubet RA, Steele VE, Kelloff GJ and Rao CV. **Chemopreventive efficacy of sulindac sulfone against colon cancer depends on time of administration during carcinogenic process.** *Cancer Res.* 1999; **59**:3387-91. | [Article](#) | [PubMed](#)
141. Harman D. **Reducing agents as chemotherapeutic agents in cancer.** *Clinical Res.* 1956; **4**:54-55.
142. Ozaslan M, Karagoz D, Kilic IH, Guldur ME. **Ehrlich ascites carcinoma.** *African J. Biotech.* 2011; **10**:2375-2378. | [Pdf](#)
143. Goodman MG and Weigle WO. **Nonspecific activation of murine lymphocytes. I. Proliferation and polyclonal activation induced by 2-mercaptoethanol and alpha-thioglycerol.** *J Exp Med.* 1977; **145**:473-89. | [Pdf](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
144. Yim CY, Hibbs JB, Jr., McGregor JR, Galinsky RE and Samlowski WE. **Use of N-acetyl cysteine to increase intracellular glutathione during the**

- induction of antitumor responses by IL-2.** *J Immunol.* 1994; **152**:5796-805. | [Article](#) | [PubMed](#)
145. Ito K, Takubo K, Arai F, Satoh H, Matsuoka S, Ohmura M, Naka K, Azuma M, Miyamoto K, Hosokawa K, Ikeda Y, Mak TW, Suda T and Hirao A. **Regulation of reactive oxygen species by Atm is essential for proper response to DNA double-strand breaks in lymphocytes.** *J Immunol.* 2007; **178**:103-10. | [Article](#) | [PubMed](#)
146. Reliene R and Schiestl RH. **Antioxidant N-acetyl cysteine reduces incidence and multiplicity of lymphoma in Atm deficient mice.** *DNA Repair (Amst).* 2006; **5**:852-9. | [Article](#) | [PubMed](#)
147. Schubert R, Erker L, Barlow C, Yakushiji H, Larson D, Russo A, Mitchell JB and Wynshaw-Boris A. **Cancer chemoprevention by the antioxidant tempol in Atm-deficient mice.** *Hum Mol Genet.* 2004; **13**:1793-802. | [Article](#) | [PubMed](#)
148. Kumagai H, Mukaisho K, Sugihara H, Miwa K, Yamamoto G and Hattori T. **Thiopropine inhibits development of esophageal adenocarcinoma induced by gastroduodenal reflux in rats.** *Carcinogenesis.* 2004; **25**:723-7. | [Article](#) | [PubMed](#)
149. Boolbol SK, Dannenberg AJ, Chadburn A, Martucci C, Guo XJ, Ramonetti JT, Abreu-Goris M, Newmark HL, Lipkin ML, DeCosse JJ and Bertagnolli MM. **Cyclooxygenase-2 overexpression and tumor formation are blocked by sulindac in a murine model of familial adenomatous polyposis.** *Cancer Res.* 1996; **56**:2556-60. | [Article](#) | [PubMed](#)
150. Mahmoud NN, Boolbol SK, Dannenberg AJ, Mestre JR, Bilinski RT, Martucci C, Newmark HL, Chadburn A and Bertagnolli MM. **The sulfide metabolite of sulindac prevents tumors and restores enterocyte apoptosis in a murine model of familial adenomatous polyposis.** *Carcinogenesis.* 1998; **19**:87-91. | [Article](#) | [PubMed](#)
151. Penzes L, Noble RC, Beregi E, Imre S, Izsak J and Regius O. **Effect of 2-mercaptoethanol on some metabolic indices of ageing of CBA/Ca inbred mice.** *Mech Ageing Dev.* 1988; **45**:75-92. | [Article](#) | [PubMed](#)
152. Aune TM and Pierce CW. **Conversion of soluble immune response suppressor to macrophage-derived suppressor factor by peroxide.** *Proc Natl Acad Sci U S A.* 1981; **78**:5099-103. | [Article](#) | [PubMed](#) | [Abstract](#) | [PubMed Full Text](#)
153. Warrington RJ, Sauder PJ, Olivier SL, Rutherford WJ and Wilkins JA. **A human T cell-derived soluble factor able to suppress pokeweed mitogen-induced immunoglobulin production.** *J Immunol.* 1983; **130**:237-41. | [Article](#) | [PubMed](#)
154. Schnaper HW, Aune TM and Pierce CW. **Suppressor T cell activation by human leukocyte interferon.** *J Immunol.* 1983; **131**:2301-6. | [Article](#) | [PubMed](#)
155. Laurence J, Gottlieb AB and Kunkel HG. **Soluble suppressor factors in patients with acquired immune deficiency syndrome and its prodrome. Elaboration in vitro by T lymphocyte-adherent cell interactions.** *J Clin Invest.* 1983; **72**:2072-81. | [Article](#) | [PubMed](#) | [Abstract](#) | [PubMed Full Text](#)
156. Redelman D and Hudig D. **The mechanism of cell-mediated cytotoxicity. I. Killing by murine cytotoxic T lymphocytes requires cell surface thiols and activated proteases.** *J Immunol.* 1980; **124**:870-8. | [Article](#) | [PubMed](#)
157. Ristow SS, Starkey JR, Stanford DR, Davis WC and Brooks CG. **Cell surface thiols, but not intracellular glutathione, are essential for cytotoxicity by a cloned murine natural killer cell line.** *Immunol Invest.* 1985; **14**:401-14. | [Article](#) | [PubMed](#)
158. Duncan DD and Lawrence DA. **Four sulfhydryl-modifying compounds cause different structural damage but similar functional damage in murine lymphocytes.** *Chem Biol Interact.* 1988; **68**:137-52. | [Article](#) | [PubMed](#)
159. Lawrence DA, Song R and Weber P. **Surface thiols of human lymphocytes and their changes after in vitro and in vivo activation.** *J Leukoc Biol.* 1996; **60**:611-8. | [Article](#) | [PubMed](#)
160. Cayota A, Vuillier F, Gonzalez G and Dighiero G. **In vitro antioxidant treatment recovers proliferative responses of anergic CD4+ lymphocytes from human immunodeficiency virus-infected individuals.** *Blood.* 1996; **87**:4746-53. | [Article](#) | [PubMed](#)
161. Hadzic T, Li L, Cheng N, Walsh SA, Spitz DR and Knudson CM. **The role of low molecular weight thiols in T lymphocyte proliferation and IL-2 secretion.** *J Immunol.* 2005; **175**:7965-72. | [Article](#) | [PubMed](#)
162. Cannizzo ES, Clement CC, Morozova K, Valdor R, Kaushik S, Almeida LN, Follo C, Sahu R, Cuervo AM, Macian F and Santambrogio L. **Age-related oxidative stress compromises endosomal proteostasis.** *Cell Rep.* 2012; **2**:136-49. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)
163. Milner JA. **Mechanisms by which garlic and allyl sulfur compounds suppress carcinogen bioactivation. Garlic and carcinogenesis.** *Adv Exp Med Biol.* 2001; **492**:69-81. | [Article](#) | [PubMed](#)
164. Wong WW, Boutros PC, Wasylshen AR, Guckert KD, O'Brien EM, Griffiths R, Martirosyan AR, Bros C, Jurisica I, Langler RF and Penn LZ. **Characterization of the apoptotic response of human leukemia cells to organosulfur compounds.** *BMC Cancer.* 2010; **10**:351. | [Article](#) | [PubMed Abstract](#) | [PubMed Full Text](#)

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